# International Journal of Pharmacognosy and Life Science

E-ISSN: 2707-2835 P-ISSN: 2707-2827

www.pharmacognosyjournal.com IJPLS 2022; 3(1): 07-14 Received: 04-11-2021 Accepted: 06-12-2021

#### IM Hassan

Department of Veterinary Physiology and Biochemistry, Usmanu Danfodiyo University Sokoto, PMB 2346, Sokoto. Sokoto, Nigeria

#### AY Abbas

Department of Biochemistry and Molecular Biology, Usmanu Danfodiyo University Sokoto, PMB 2346, Sokoto, Nigeria

#### SA Balarabe

Department of Internal Medicine, Usmanu Danfodiyo University Sokoto, PMB 2346, Sokoto, Nigeria

#### Y Saidu

Department of Biochemistry and Molecular Biology, Usmanu Danfodiyo University Sokoto, PMB 2346, Sokoto, Nigeria

#### LS Bilbis

Department of Biochemistry and Molecular Biology, Usmanu Danfodiyo University Sokoto, PMB 2346, Sokoto, Nigeria

### Corresponding Author: IM Hassan

Department of Veterinary Physiology and Biochemistry, Usmanu Danfodiyo University Sokoto, PMB 2346, Sokoto. Sokoto, Nigeria

## Evaluation of anti-oxidant, anti-cholinesterase activity and dopamine levels of methanolic leaves extract of Solanum incanum in arsenic-induced neurodegeneration in Wistar rats

IM Hassan, AY Abbas, SA Balarabe, Y Saidu and LS Bilbis

**DOI:** https://doi.org/10.33545/27072827.2022.v3.i1a.39

#### Abstract

The study investigated the anti-oxidant, anti-cholinesterase activity and dopamine levels of methanolic leaves extract Solanum incanum in arsenic-induced neurodegeneration. Animals were divided into four groups: A Control (maintained on food and water only), B treated with 300 mg/kg S. incanum, C treated with 20 mg/kg As<sub>2</sub>O<sub>3</sub>, and D treated with 300 mg/kg and then 20 mg/kg As<sub>2</sub>O<sub>3</sub>. Serum antioxidant activities were determined by measuring the Catalase (CAT) and Super-oxidase (SOD) activities using enzyme-linked immunosorbent assay (ELISA). Serum acetylcholinesterase inhibitory activity and dopamine levels were also quantified using ELISA. Prepared slides from the brain were dehydrated using a dry air oven in xylene for 30 minutes and later mounted on the microscope and viewed using oil immersion ×1000 magnification. The result of CAT and SOD assays showed significant decrease (P<0.0002) when control groups (40 µmol/min/mL and 107.7 U/mL) were compared with the group exposed to arsenic only (20 µmol/min/mL and 71.32 U/mL) respectively. The activity of cholinesterase was significantly reduced (P<0.0006) in group exposed to arsenic only (30.30 U/mL) compared to the control group (45.42 U/mL) But, there was no significant difference between the control group and group treated with S. incanum methanol extract and then arsenic and group treated with arsenic and then S. incanum methanol extract. There was no observed significant difference (P>0.5) in dopamine levels between the groups. Histopathological lesions observed include degenerative vacuolated neurocytes. For histopathological study, there were dystrophic changes in the form of shrunken hyperchromatic neurocytes which were irregular with chromatolysis and abnormal Nissl granule distribution in groups exposed to arsenic only as well as arsenic S. incanum. Because of the presence of significant anti-oxidant and anticholinesterase activities as well as neuronal cells protective effect, S. incanum is a potential source of remedy against arsenic poisoning.

**Keywords:** Antioxidant activity, cholinesterase inhibitors, crude extract, dopamine activity, oxidative, toxicity study

#### Introduction

Nerve cells in the brain and peripheral nervous system lose function over time and eventually die in neurodegenerative disease (Borah *et al.*, 2021) <sup>[7]</sup>. Although therapies may alleviate some of the physical or mental symptoms associated with neurodegenerative disorders, there are presently no known cures or ways to decrease disease development (Lee *et al.*, 2019) <sup>[16]</sup>. Scientists have discovered that a person's genes and environment factors increase the chance of acquiring a neurodegeneration (Scheiblich *et al.*, 2020) <sup>[33]</sup>. Chronic arsenic exposure has been related to a variety of negative health impacts in exposed individuals, including skin disorders, cancer, diabetes, cardiovascular disease, reproductive, developmental, and neurodegeneration (Mostafalou and Abdollahi, 2013) <sup>[23]</sup>.

The biological properties of *Solanum incanum* or garden egg leaf and seed as well as its derivatives have been reported, including antioxidant neuroprotectant properties (Patel *et al.*, 2021) <sup>[28]</sup>. Studies shows that garden egg is used in the treatment of variety of diseases conditions such as diabetic retinopathy, uveitis, light induced retinopathy, and ischemia/reperfusion injury (Yang *et al.*, 2019) <sup>[45]</sup>. The plant is said to contain important minerals and vitamins such as proteins, riboflavin, thiamine iron, calcium, nicotinamide, glucose, Vitamin C water, fiber, fat, and carotenes, all of which are beneficial to one's health (Abdisa, 2019) <sup>[1]</sup>. Carotene is converted to Vitamin A in the body, which is required for

night blindness prevention and xerophthalmia (Gomes *et al.*, 2013) [10]. The high concentration of  $\alpha$ -chaconine in this plant have anticholinesterase action in nerve tissues, which contributes to its therapeutic usefulness (Väänänen *et al.*, 2005) [41].

Cholinesterase inhibitors work by preventing cholinesterase from hydrolyzing acetylcholine into its constituent's acetate and choline. This increases acetylcholine availability and duration of action in neuromuscular junctions (Hassan et al., 2020) [12]. There are two active sites in the cholinesterase enzyme: an anionic site created by tryptophan and an esteractic site formed by serine (Sharma, 2019) [36]. Organophosphates and arsenic, for example, interact with the serine esteractic site of cholinesterase, preventing it from cleaving acetylcholine. As a consequence, acetylcholine will continue to build up and activate receptors in the brain (Colovic et al., 2013) [9]. Cholinesterase inhibitors are divided into three categories: reversible, irreversible, and pseudo-reversible (Strelnik et al., 2016) [38]. In general, reversible cholinesterase inhibitors are used for therapeutic purposes. Irreversible and pseudo-reversible inhibitors, on the other hand, are often employed as insecticides and biowarfare (nerve agents) (Colovic et al., 2013) [9].

Despite the growing popularity of *Solanum incanum* leaf and seed as a supplements and or therapeutic agents, little is known about its antioxidant and anti-cholinesterase effects. As a result, the present study evaluated the effect of methanol leaf extract of *Solanum incanum* on antioxidant enzymes, cholinesterase inhibition and histopathologic effect in the brain of Wistar rats.

#### Materials and Method

#### **Plants Collection and Identification**

Solanum incanum leaf was obtained from Gwaski village, Sakwa ward, Hawul LGA, Borno State, Nigeria. S. Sanusi from the University of Maiduguri's Department of Biological Science in Borno State, Nigeria, identified it, and a voucher number (DCPT 014) was assigned. The leaf was cleaned and air-dried for two weeks at ambient temperature  $(26 \pm 1~^{\circ}\text{C})$ .

#### **Plant Extraction**

Solanum incanum leaves were washed and crushed to semipowdered form (40–60 mesh). The powder was allowed to dry for two weeks at room temperature (26  $\pm$  1 °C). In flat bottom flasks (Sigma-Aldrich, USA), 200 g of leaves sample were soaked for 3 days in 1000 mL of 80 percent methanol. To get a high yield of the extract, the semipowdered leaves in methanol was shaken daily for three days at 25  $\pm$  1 °C. The extract was then filtered using new white clean muslin cloth and concentrated to semisolid form using a rotary evaporator (IKA® RV 10, USA) at 42 °C. The semi-solid crude extract was then weighed and put into sample vials, which were then kept at 4 °C until needed. Yield (%) = [wt of extract (g)/wt of plant material (g)] × 100 (Hassan *et al.*, 2020) [12].

#### **Plants Sample Dilution and Dose Preparation**

Stock solution was prepared by dissolving 100 g of *S. incanum* leaves extract in 1 L of 100% DMSO (100 g/L). Preparation of sub-stocks solution was done by diluting the stock solution to 10 mg/mL with distilled water. Working solution was prepared from sub-stock solution using twofold serial dilution with distilled water at concentrations of

interest (1 mg/mL). DMSO (vehicle) was maintained at 0.1% in all concentration of extract (Hassan *et al.*, 2020) [12].

#### Toxicity study of the extract

Oral toxicity study of the extract) was carried out in accordance with the Organization for Economic Cooperation and Development's guidelines (OECD) 423(7) (Alli et al., 2011) [4]. The Usmanu Danfodiyo University Institutional Animal Care and use Committee (IACUC) Faculty of Veterinary Medicine Usmanu Danfodiyo University Sokoto, Nigeria approved the protocol for these under number UDUS/IACUC/AUPexperiments R005/2020). For the acute toxicity study of the crude extract, rats were treated with 1000, 500 and 250 mg/kg of the extract for 2 days (Clemente et al., 2019) [8]. Probit analysis was used to determine the LC<sub>50</sub>.In chronic toxicity study of the crude extract, 500, 250 and 125 mg/kg of the extract were administered for 14 days (Adekola et al., 2020)

#### **Induction of Neurodegeneration using Arsenic**

Neurodegeneration in in rats was induced by administering arsenic trioxide ( $As_2O_3$ ) (20 mg/kg bw) orally for 5 days (Patlolla and Tchounwou, 2005) [29].

# Protective and therapeutic effects of the extract on arsenic-induced Neurodegeneration

Neuroprotective and therapeutic effects of the extract on arsenic-induced neurodegeneration were carried out in accordance with the OECD guidelines. Animals were divided into four groups of 5 rats each. The groups were treated as follows:

- A. Normal control, received only food and water.
- B. In addition to food and water, the rats were administered 300 mg/kg bw of *S. incanum* methanol extract.
- C. Rats received 20 mg/kg bw of arsenic trioxide (As<sub>2</sub>O<sub>3</sub>) in addition to food and water.
- D. Rats were given 300 mg/kg bw of *S. incanum* methanol extract for 10 days followed by administration of 20 mg/kg bw of arsenic trioxide ( $As_2O_3$ ) for 5 days in addition to food and water.

degenerative vacuolated neurocytes degenerative vacuolated neurocytes *S. incanum* methanol extract E. Rats were given 20 mg/kg bw of arsenic trioxide (As<sub>2</sub>O<sub>3</sub>) for 5 days followed by administration 300 mg/kg bw of *S. incanum* methanol extract for 10 days in addition to food and water.

Animals were anaesthetized with chloroform vapor, fortyeight (48) hours after the last treatment, blood was collected via cardiac puncture with 5 ml syringe and needle and transferred into EDTA free bottles. The brain of each rat was removed after the head was dissected with the use of a dissecting kit. Before putting into a clean sample container containing 10% neutral buffered formalin, the dissected brains were dipped into a beaker containing normal saline to wipe away excess blood (Parasuraman *et al.*, 2010) [27].

#### **Biochemical Tests**

Total proteins (Kashyap *et al.*, 2020) <sup>[14]</sup>, Catalase (CAT) activity (Zhang *et al.*, 2017) <sup>[46]</sup>, Super-oxidase (SOD) activity (Warsinggih *et al.*, 2020) <sup>[44]</sup>, Cholinesterase (Ache) inhibitory activity (Hadda *et al.*, 2017) <sup>[11]</sup> and Dopamine (DA) levels (van Nie *et al.*, 2020) were determined from serum using enzyme-linked immunosorbent assay (ELISA)

assay kits from Thermo Fisher Scientific, USA as described in the manufacturer's guide.

#### **Histological Study**

The brain was immersed in fixative (10% neutral buffered formalin) for 3 days and then transferred directly to 70% alcohol where it was graded to 90%,100% alcohol for 8, 12 and 15 hours respectively. Alcohol was then replaced with Xylene and incubated 4 hours, followed by embedding the tissues and insertion into paraffin wax to harden tissue for easy cutting into thin sections with the microtome. Casting of tissues into block of paraffin in 'L' block was carried out to remove air bubbles followed by it solidification. Prepared slides were dehydrated using a dry air oven in xylene for 30 minutes and latter mounted on the microscope and viewed using oil immersion ×1000 magnification (Sabdyusheva *et al.*, 2020) [31].

#### **Results**

#### **Toxicity study**

The chronic toxicity studies revealed the  $LC_{50}$  of 676.10 mg/Kg body weight for the extract and 45.70 mg/Kg body weight for the arsenic trioxide (As<sub>2</sub>O<sub>3</sub>).

#### **Total Protein**

Result of total protein shows protein level of 7.47g/dL in the control group compared to the group exposed to arsenic only. There is significant decrease in protein level at P<0.01 in the group that was exposed to arsenic (As<sub>2</sub>O<sub>3</sub>) only. (Figure 1).

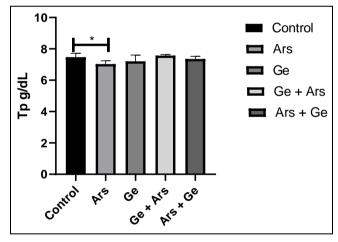
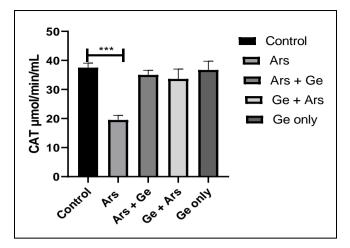


Fig 1: Serum Total protein (TP) of Wistar rats treated with arsenic (Ars), *S. incanum* methanol extract (Ge), *S. incanum* methanol extract (Ge) and then arsenic (Ars), arsenic (Ars) and then *S. incanum* (Ge). \*P<0.01 represents significant different between the control group and group exposed to arsenic (Ars) only. The values represent mean ± SEM from independent experiments.

#### Catalase (CAT) Activity

Result of catalase (CAT) activity showed and with significant different (P<0.0002) between the control group (40 µmol/min/mL) and group exposed to arsenic (20 µmol/min/mL) only. There was no significant difference between the control group and groups treated with crude extract, crude extract and arsenic (As<sub>2</sub>O<sub>3</sub>), arsenic (As<sub>2</sub>O<sub>3</sub>) and then crude extract (Figure 2).



**Fig 2:** Catalase (CAT) activity of Wistar rats treated with arsenic (Ars), *S. incanum* methanol *extract* (Ge), *S. incanum* methanol extract (Ge) and then arsenic (Ars), arsenic (Ars) and then *S. incanum* methanol extract (Ge). \*\*\*\*P<0.0002 represents significant different values of the control group and group exposed to arsenic (Ars) only. The values represent mean ± SEM from independent experiments.

#### Superoxidase (SOD) Activity

Result of superoxidase (SOD) activity showed and with significant different (P<0.0048) between the control group (107.7 U/mL) and group exposed to arsenic (71.32 U/mL) only. There was no significant difference between the control group and groups treated with crude extract, crude extract and then arsenic ( $As_2O_3$ ), arsenic ( $As_2O_3$ ) and then crude extract (Figure 3).

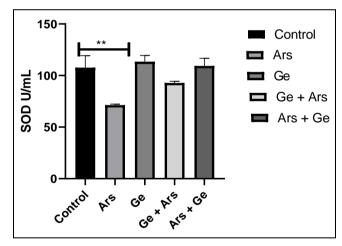


Fig 3: Superoxidase (SOD) activity of Wistar treated with arsenic (Ars), *S. incanum* methanol extract (Ge) and arsenic (Ars), arsenic (Ars) and *S. incanum* methanol extract (Ge). \*\*P<0.0048 represents significant different values of the control group and group exposed to arsenic (Ars) only. The values represent mean ± SEM from independent experiments.

#### Cholinesterase (AChE) Inhibitory Activity

Result of cholinesterase (AChE) inhibitory activity showed and with significant difference (*P*<0.0006) between the control group (30.30 U/mL) and group exposed to arsenic (45.42 U/mL) only. There was no significant difference between the control group and groups treated with *S. incanum* methanol extract, *S. incanum* methanol extract and then arsenic (Ars), arsenic (Ars) and then *S. incanum* methanol extract (Figure 4).

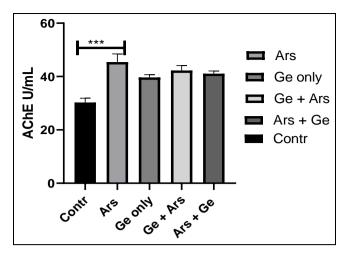


Fig 4: Acetyl cholinesterase (AChE) assay of Wistar treated with arsenic (Ars), *S. incanum* methanol extract (Ge), *S. incanum* methanol extract (Ge) and arsenic (Ars), arsenic (Ars) and *S. incanum* methanol extract (Ge).\*\*\*P<0.0006 represents significant different values of the control group and group exposed to arsenic (As<sub>2</sub>O<sub>3</sub>) only. The values represent mean  $\pm$  SEM from independent experiments.

#### Dopamine (DA) Levels

There was no observed significant different (P > 0.5) in dopamine levels between the groups. The values obtained were 128.3 U/mL (control), 121.7 U/mL (arsenic), 113.2 U/mL (*S. incanum* methanol extract), 131.2 U/mL (*S. incanum* methanol extract + arsenic) 123.4 U/mL (arsenic + *S. incanum* methanol extract). (Figure 5).

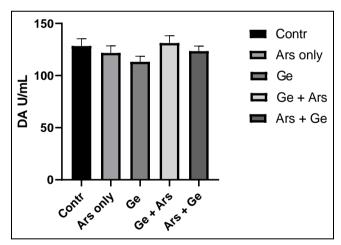


Fig 5: Dopamine (DA) levels from Wistar treated with arsenic (Ars), *S. incanum* methanol extract, *S. incanum* methanol extract and arsenic (Ars), arsenic (Ars) and *S. incanum* methanol extract). There was no significant different (P > 0.5) between the control groups and groups exposed to arsenic (Ars), *S. incanum* methanol extract, *S. incanum* methanol extract and arsenic (Ars), arsenic (Ars) and *S. incanum* methanol extract. The values represent mean  $\pm$  SEM from independent experiments.

#### Histopathologic effect on brain

Results of histopathological finding from Wistar rats showed lesions such as degenerative vacuolated neurocytes, dystrophic changes in the form of shrunken hyperchromatic, irregular with chromatolysis in neurons and abnormal Nissl granule distribution in group exposed to arsenic (Ars) only (plate D).

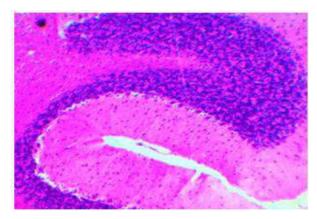
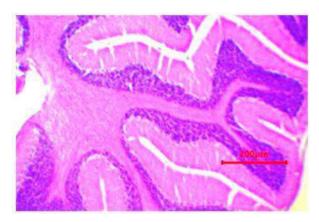
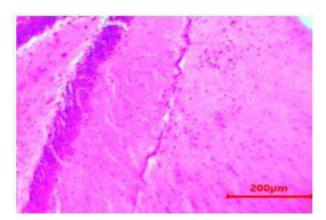


Plate A: Brain section of the Wistar rat fed with feed and water only (control group)



**Plate B:** Brain section of the Wistar rat exposed to *S. incanum* methanol extract only



**Plate C:** Brain section of the Wistar rat exposed to *S. incanum* methanol extract and then arsenic (Ars)

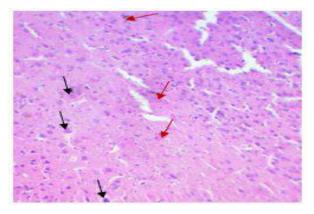
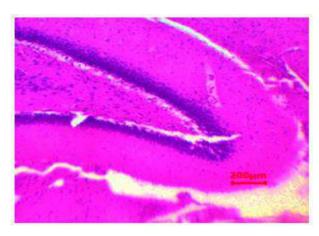


Plate D: Brain section of the Wistar rat exposed to arsenic (Ars) only and Plate



**Plate** E: Brain section of the Wistar rat exposed to arsenic (Ars) and then *S. incanum* methanol extract.

**Fig 6:** Histopathological assessment of the brain of Wistar rats: The brain showed degenerative vacuolated neurocytes with red dot (red arrows), dystrophic changes in the form of shrunken hyperchromatic, irregular with chromatolysis in neurons and abnormal Nissl granule distribution with blue dot (black arrows)

#### **Discussion**

The main purpose of this study was to determine the efficacy of *Solanum incanum* in protecting and ameliorating the neurodegenerative effect caused by arsenic trioxde  $(As_2O_3)$  in Wistar albino rats.

Effect of arsenic trioxide on animals' serum total protein has been studied by many researchers (Sun et al., 2021. Mingxing et al., 2019). Groups that were exposed to arsenic trioxide (As<sub>2</sub>O<sub>3</sub>) only showed significant decrease in total protein (p<0.01) compared to the control and other treated groups. On the effect of Solanum incanum on serum protein, there was little research. Some studies have found that Solanum incanum methanol leaf extract can protect serum protein from inflammatory agents by inhibiting cells from activating and releasing inflammatory mediators that promote vasodilation and increased blood permeability. This also prevents plasma proteins and fluids from leaking into the tissues (Anosike et al., 2012) [5]. This may be as a result of protein denaturation by the arsenic trioxide (As<sub>2</sub>O<sub>3</sub>). Sulfhydryl groups have a strong affinity for trivalent arsenicals, and they can bind to destroy cysteines in peptides and proteins (Shen et al., 2013) [37]. This is in contrary to the finding by Herrera et al., (2021) [13], who reported increased in total protein (TP,) after arsenic administration in albino rats, leading to the damage of cell and followed by acute inflammation marked by significant neutrophil infiltration.

The serum antioxidant enzymes CAT and SOD were examined in this study and the findings demonstrated a significant decrease in both CAT and SOD (P<0.0002, P<0.0048) in Arsenic-treated rats compared to the control group. Antioxidant effect of *Solanum incanum* leave extract may be due to the present of several bioactive compounds. Present of bioactive such as phenol, flavonoid, beta carotene and ascorbic acid proved the antioxidant properties of plant extract (Adelakun *et al.*, 2020) [3]. Phenolic compounds work by removing free radicals, binding metal ions, inhibiting enzymatic systems that produce free radicals, increasing the concentration of biologically important endogenous antioxidants, and inducing the expression of a variety of genes involved in the synthesis of enzymes that protect against oxidative stress (Valko, Rhodes, Moncol,

Izakovic, & Mazur, 2006) [42]. Many flavonoids have been shown to have considerable antioxidant activity in diverse in vitro systems (Kaurinovic & Vastag, 2019) [15]. This is in line with the finding reported by Seif *et al.*, (2021) [34] on the anti-oxidative stress effect of *Zingiber officinale* ethanolic extract in in male rats exposed to arsenic. Antioxidant enzymes aid in the fight against free radicals/oxygenderived species produced during normal physiological processes (Liu *et al.*, 2020) [17].

S. incanum methanol extract was tested for it cholinesterase inhibitory activity and effect on dopamine levels. The results obtained showed that there was a significant reduction in cholinesterase enzyme activity (P<0.0006) in the groups treated with arsenic trioxide (Ars) only. There was no observed statistical different in dopamine levels between the groups that were exposed to arsenic trioxide (Ars) only and the other groups. There was no available material on anticholinesterase effects Solanum incanum's. Lucky et al., (2018) demonstrated the ability of a secondary metabolite from Moringa oleifera extract to neutralize free radicals, preventing the formation of senile plaques (A) and neurofibrillary tangles (NFTs) (tau protein) in the hippocampus and cerebral cortex of neurodegenerative induced rats. Neuronal injury and synaptic dysfunction are caused by protein aggregates (A and tau proteins). Several medicinal plants have been shown to have anticholinesterase inhibitory activities in the literature (Mathew & Subramanian, 2014 [19], Owokotomo, Ekundayo et al., 2015 [26], Samaradivakara et al., 2016 [32], Malar et al., 2017 [18], Ovais et al., 2018) [25]. Furthermore, antioxidants such as vitamin E and vitamin C have been linked to a decreased incidence and prevalence of Alzheimer's disease(AD) and AD patients taking large doses of antioxidants have been shown to have a slower pace of cognitive decline (Polidori & Nelles, 2014) [30]. There is high tendency of correlating the anticholinesterase and antioxidant effect of this plant with it phytochemical content. AChE inhibitory activity has been discovered in a wide range of plants. Alkaloids, ursolic acid, lignans, flavonoids, terpenoids, and coumarins are phytochemicals that may be responsible for this activity (Mottay & Neergheen-Bhujun, 2016) [24]. Result of dopamine in this study contradicts that of Moreno et al., (2016) who discovered arsenic exposure resulted in hypoactivity at six months due to increase in dopamine levels. The non-statistical difference observed in this study may be as a result of short exposure period (10 days). Several literatures documented arsenic to have neurotoxic effects induces increase secretion of dopamine (DA) and serotonin (5-HT) due to regulation of norepinephrine (NE) levels (Thakur et al., 2021) [40].

Histopathological study also showed degenerative vacuolated neurocytes, dystrophic changes in the form of shrunken hyperchromatic, irregular with chromatolysis in neurons and abnormal Nissl granule distribution in group exposed to arsenic (Ars) only plate D. The histological lesions may be due to the toxic effect of arsenic on the neuronal cells leading to degenerative changes and death of neurons. There was no observed lesion in the control group, group treated with S. incanum methanol leave extract only, group treated with S. incanum methanol leave extract for 5 days and then exposed to arsenic and group that were exposed to arsenic 5 days before treatment with S. incanum methanol leave extract for 10 days. This result is similar with the finding reported by Shaibah et al., 2016) [35] and

differed with the finding reported by Mohammod, (2015) [21] who showed edema, intracellular space, edematous changes in arsenic exposed brain tissue of mice. The result also differed with that of Bashir *et al.*, (2006) [6], who reported cellular necrosis due to nuclear pyknosis following 24 hours mice exposure to arsenic. Dissimilarity may be due to specie difference. Mohammod and Bashir *et al.*, carried out their research on mice while our on study was conducted on Wistar rat.

#### Conclusions

This study showed that the *S. incanum* methanol extract have significant anti-oxidant and anti-cholinesterase activity. Histopathologic result revealed the protective and curative effect of this extract on arsenic induces neurodegeneration. Because of the presence of significant anti-oxidant, anticholinesterases activity as well as neuronal cells protective and curative effect of this plant, it could be a potential source of lead compounds for the development of drugs that can be used in the management of neurodegenerative diseases.

#### **Data Availability**

Data are available from Dr Ibrahim Maina Hassan but with the permission of Usmanu Danfodiyo University Sokoto and TETFund.

#### **Conflicts of Interest**

The authors declare no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

#### Acknowledgments

The authors would like to thank Usmanu Danfodiyo University and Tertiary Education Trust Fund (TET fund) for their support and sponsorship through Institution Based Research (IBR). The authors wish to acknowledge the efforts of Dr. Abdullahi Abdullahi Raji of the Department of Veterinary Pathology and Oncology, Usmanu Danfodiyo University Sokoto, for the reading and interpretation of the histopathological slides.

#### References

- Abdisa T. Medicinal Value of Croton macrostachyus and Solanum incanum against Causative Agent of Foodborne Diseases. Veterinary Medicine – Open Journal. 2019. https://doi.org/10.17140/vmoj-4-137
- 2. Adekola MB, Areola JO, Omisore NO, Asaolu FT, Ogunleye SG, Apalowo OE *et al.* Sub-chronic toxicity study of ethanol stem-bark extract of *Blighia sapida* (Sapindaceae) in wistar rats. Heliyon. 2020. https://doi.org/10.1016/j.heliyon.2019.e02801
- 3. Adelakun SA, Ukwenya VO, Akingbade GT, Omotoso OD, Aniah JA. Interventions of aqueous extract of Solanum melongena fruits (garden eggs) on mercury chloride induced testicular toxicity in adult male Wistar rats. Biomedical Journal. 2020. https://doi.org/10.1016/j.bj.2019.07.004
- 4. Alli LA, Adesokan AA, Salawu AO, Akanji MA, & Tijani AY. Anti-plasmodial activity of aqueous root extract of *Acacia nilotica*. African Journal of Biochemistry Research. 2011.
- Anosike CA, Obidoa O, Ezeanyika LU. Membrane stabilization as a mechanism of the anti-inflammatory

- activity of methanol extract of garden egg (*Solanum aethiopicum*). DARU, Journal of Pharmaceutical Sciences. 2012. https://doi.org/10.1186/2008-2231-20-76
- Bashir S, Sharma Y, Irshad M, Gupta SD, Dogra TD. Arsenic-induced cell death in liver and brain of experimental rats. Basic and Clinical Pharmacology and Toxicology. 2006. https://doi.org/10.1111/j.1742-7843.2006.pto\_170.x
- Borah P, Deb PK, Chandrasekaran B, Goyal M, Bansal M, Hussain S, Singh V. Neurological Consequences of SARS-CoV-2 Infection and Concurrence of Treatment-Induced Neuropsychiatric Adverse Events in COVID-19 Patients: Navigating the Uncharted. Frontiers in Molecular Biosciences. 2021. https://doi.org/10.3389/fmolb.2021.627723
- Clemente M, Miguel MD, Felipe KB, Gribner C, Moura PF, Rigoni AGR, et al. Acute and sub-acute oral toxicity studies of standardized extract of *Nasturtium* officinale in Wistar rats. Regulatory Toxicology and Pharmacology. 2019. https://doi.org/10.1016/j.yrtph.2019.104443
- Colovic MB, Krstic DZ, Lazarevic-Pasti TD, Bondzic AM, Vasic VM. Acetylcholinesterase Inhibitors: Pharmacology and Toxicology. Current Neuropharmacology. 2013. https://doi.org/10.2174/1570159x11311030006
- 10. Gomes Ribeiro EM, Jaegar de Carvalho LM, de Azevedo Sarmet Smiderle L, Dellamora Ortiz GM. Chapter III: Vitamin A: Dietary Sources and Health Consequences. Vitamin A and Vitamin E: Daily Requirements, Dietary Sources and Symptoms of Deficiency, 2013.
- 11. Hadda T Ben, Talhi O, Silva ASM, Senol FS, Orhan I E, Rauf A Mubarak MS. Cholinesterase Inhibitory Activity of Some semi-Rigid Spiro Heterocycles: POM analyses and Crystalline Structure of Pharmacophore Site. Mini-Reviews in Medicinal Chemistry, 2017. https://doi.org/10.2174/1389557517666170713114039
- 12. Hassan I, Wan Ibrahim WN, Yusuf FM, Ahmad SA, Ahmad S. Biochemical Constituent of Ginkgo biloba (Seed) 80% Methanol Extract Inhibits Cholinesterase Enzymes in Javanese Medaka (*Oryzias javanicus*) Model. Journal of Toxicology. 2020. https://doi.org/10.1155/2020/8815313
- Herrera AS, Beeraka NM, Sinelnikov MY, Nikolenko VN, Giller DB, Solis LFT. The Beneficial Effects of QIAPI 1® against Pentavalent Arsenic-Induced Lung Toxicity a Hypothetical Model for SARS CoV2-Induced Lung Toxicity. Current Pharmaceutical Biotechnology. 2021. https://doi.org/10.2174/1389201022666210412142230
- 14. Kashyap SP, Hiremath J, Vinutha S, Patil SS, Suresh KP, Roy P, *et al.* Development of recombinant nucleocapsid protein-based indirect enzyme-linked immunosorbent assay for sero-survey of porcine reproductive and respiratory syndrome. Veterinary World. 2020.
  - https://doi.org/10.14202/VETWORLD.2020.2587-2595
- Kaurinovic B, Vastag D. Flavonoids and Phenolic Acids as Potential Natural Antioxidants. In Antioxidants. 2019. https://doi.org/10.5772/intechopen.83731

- 16. Lee G, Cummings J, Decourt B, Leverenz JB, Sabbagh MN. Clinical drug development for dementia with Lewy bodies: past and present. Expert Opinion on Investigational Drugs. 2019. https://doi.org/10.1080/13543784.2019.1681398
- 17. Liu Y, Liang Y, Zheng B, Chu L, Ma D, Wang H, et al. Protective effects of crocetin on arsenic trioxideinduced hepatic injury: Involvement of suppression in oxidative stress and inflammation through activation of nrf2 signaling pathway in rats. Drug Design, Development and Therapy, 2020. https://doi.org/10.2147/DDDT.S247947
- 18. Malar DS, Shafreen RB, Pandian SK, Devi KP. Cholinesterase inhibitory, anti-amyloidogenic neuroprotective effect of the medicinal plant Grewia tiliaefolia - An in vitro and in silico study. Pharmaceutical Biology. 2017. https://doi.org/10.1080/13880209.2016.1241811
- 19. Mathew M, Subramanian S. In vitro screening for anticholinesterase and antioxidant activity of methanolic extracts of ayurvedic medicinal plants used for cognitive disorders. **PLoS** ONE. 2014. https://doi.org/10.1371/journal.pone.0086804
- 20. Mingxing S, Haiying W, Congsong S, Chunyu Y, Liu C, Wang Q. Acute toxicity of intratracheal arsenic trioxide instillation in rat lungs. Journal of Applied Toxicology. 2019. https://doi.org/10.1002/jat.3841
- Arsenic-induced 21. Mohammod AS. Histological Alterations in Various Organs of Mice. Journal of Cytology & Histology. 2015. https://doi.org/10.4172/2157-7099.1000323
- 22. HA Ibekwe. *In vitro* anthelmintic activities of aqueous extract of Azadirachta indica Paramphistomum cervi and Fasciola hepatica. Int J Vet Sci Anim Husbandry 2019;4(1):14-18. https://doi.org/10.1155/2016/4763434
- 23. Mostafalou S, Abdollahi M. Pesticides and human chronic diseases: Evidences, mechanisms, perspectives. Toxicology and Applied Pharmacology. 2013. https://doi.org/10.1016/j.taap.2013.01.025
- 24. Mottay D, Neergheen-Bhujun VS. Anticholinesterase and Antioxidant Effects of Traditional Medicines used in the Management Neurodegenerative Diseases in Mauritius. Archives of Medical and Biomedical Research. 2016. https://doi.org/10.4314/ambr.v2i4.2
- 25. Ovais M, Ayaz M, Khalil AT, Shah SA, Jan MS, Raza A, Shinwari ZK. HPLC-DAD finger printing, antioxidant. cholinesterase, and α-glucosidase inhibitory potentials of a novel plant Olax nana. BMC Complementary and Alternative Medicine. 2018. https://doi.org/10.1186/s12906-017-2057-9
- 26. Owokotomo IA, Ekundayo O, Abayomi TG, Chukwuka AV. In-vitro anti-cholinesterase activity of essential oil from four tropical medicinal plants. Toxicology Reports. 2015. https://doi.org/10.1016/j.toxrep.2015.05.003
- 27. Parasuraman S, Raveendran R, Kesavan R. Blood sample collection in small laboratory animals. Journal of Pharmacology and Pharmacotherapeutics. 2010. https://doi.org/10.4103/0976-500X.72350
- 28. Patel P, Prasad A, Srivastava K, Singh SS, Chakrabarty D, Misra P. Updates on steroidal alkaloids and glycoalkaloids in Solanum spp.: Biosynthesis, in vitro

- production and pharmacological values. In Studies in Products Chemistry. Natural https://doi.org/10.1016/B978-0-12-819487-4.00012-4
- 29. Patlolla AK, Tchounwou PB. Cytogenetic evaluation of arsenic trioxide toxicity in Sprague-Dawley rats. Mutation Research - Genetic Toxicology Environmental Mutagenesis, 2005. https://doi.org/10.1016/j.mrgentox.2005.08.007
- 30. Polidori M, Nelles G. Antioxidant Clinical Trials in Mild Cognitive Impairment and Alzheimer's disease -Challenges and Perspectives. Current Pharmaceutical Design, 2014. https://doi.org/10.2174/13816128113196660706
- 31. Sabdyusheva Litschauer I, Becker K, Saghafi S, Ballke S, Bollwein C, Foroughipour M, et al. 3D histopathology of human tumours by fast clearing and ultramicroscopy. Scientific Reports, https://doi.org/10.1038/s41598-020-71737-w
- 32. Samaradiyakara SP, Samarasekera R, Handunnetti SM, Weerasena OVDSJ. Cholinesterase, protease inhibitory and antioxidant capacities of Sri Lankan medicinal plants. Industrial Crops and Products, https://doi.org/10.1016/j.indcrop.2015.12.047
- 33. Scheiblich H, Trombly M, Ramirez A, Heneka MT. Neuroimmune Connections in Aging Neurodegenerative Diseases. Trends in Immunology. 2020. https://doi.org/10.1016/j.it.2020.02.002
- 34. Seif M, Abd El-Aziz T, Sayed M, Wang Z. Zingiber officinale ethanolic extract attenuates oxidative stress, steroidogenic gene expression alterations, and testicular histopathology induced by sodium arsenite in male rats. Environmental Science and Pollution Research. 2021. https://doi.org/10.1007/s11356-020-11509-1
- 35. Shaibah H, Elsify AE, Medhat T, Rezk H, El-Sherbiny M. Histopathological and immunohistochemical study of the protective effect of triptorelin on the neurocytes of the hippocampus and the cerebral cortex of male rats after short-term exposure cyclophosphamide. Journal of Microscopy Ultrastructure. 2016. https://doi.org/10.1016/j.jmau.2015.12.002
- 36. Sharma K. Cholinesterase inhibitors as Alzheimer's therapeutics (Review). Molecular Medicine Reports. 2019. https://doi.org/10.3892/mmr.2019.10374
- 37. Shen S, Li XF, Cullen WR, Weinfeld M, Le XC. Arsenic binding to proteins. Chemical Reviews, 2013. https://doi.org/10.1021/cr300015c
- 38. Strelnik AD, Petukhov AS, Zueva IV, Zobov VV, Petrov KA, Nikolsky EE, et al. Novel potent pyridoxine-based inhibitors of AChE and BChE, structural analogs of pyridostigmine, with improved in vivo safety profile. Bioorganic and Medicinal Chemistry Letters, 2016.
  - https://doi.org/10.1016/j.bmcl.2016.06.070
- 39. Sun Z, Cao Y, Xing Y, Wu M, Shao X, Huang Q, et al. Antiangiogenic effect of arsenic trioxide in HUVECs FoxO3a-regulated autophagy. Journal of Biochemical and Molecular Toxicology, https://doi.org/10.1002/jbt.22728
- 40. Thakur M, Rachamalla M, Niyogi S, Datusalia AK, Flora SJS. Molecular mechanism of arsenic-induced including dysfunctions. neurotoxicity neuronal International Journal of Molecular Sciences, 2021. https://doi.org/10.3390/ijms221810077

- Väänänen T, Ikonen T, Rokka VM, Kuronen P, Serimaa R, Ollilainen V. Influence of incorporated wild Solanum genomes on potato properties in terms of starch nanostructure and glycoalkaloid content. Journal of Agricultural and Food Chemistry, 2005. https://doi.org/10.1021/jf0501342
- 42. Valko M, Rhodes CJ, Moncol J, Izakovic M, Mazur M. Free radicals, metals and antioxidants in oxidative stress-induced cancer. Chemico-Biological Interactions, 2006. https://doi.org/10.1016/j.cbi.2005.12.009
- 43. Van Nie L, Salinas-Tejedor L, Dychus N, Fasbender F, Hülser ML, Cutolo M, Capellino S. Dopamine induces in vitro migration of synovial fibroblast from patients with rheumatoid arthritis. Scientific Reports, 2020. https://doi.org/10.1038/s41598-020-68836-z
- 44. Warsinggih Irawan B, Labeda I, Lusikooy RE, Sampetoding S, Kusuma MI, Faruk M. Association of superoxide dismutase enzyme with staging and grade of differentiation colorectal cancer: A cross-sectional study. Annals of Medicine and Surgery, 2020. https://doi.org/10.1016/j.amsu.2020.08.032
- 45. Yang L, Ren S, Xu F, Ma Z, Liu X, Wang L. Recent Advances in the Pharmacological Activities of Dioscin. BioMed Research International, 2019. https://doi.org/10.1155/2019/5763602
- 46. Zhang J, Chen R, Yu Z, Xue L. Superoxide Dismutase (SOD) and Catalase (CAT) Activity Assay Protocols for Caenorhabditis elegans. BIO-PROTOCOL, 2017. https://doi.org/10.21769/bioprotoc.2505.
- 47. Moreno Ávila CL, Limón-Pacheco JH, Giordano M, Rodríguez VM. Chronic Exposure to Arsenic in Drinking Water Causes Alterations in Locomotor Activity and Decreases Striatal mRNA for the D2 Dopamine Receptor in CD1 Male Mice. Journal of Toxicology. 2016.