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Ameliorative effect of *Emblica officinalis* (Amla) on altered biochemical markers and oxidative stress indices after sub-acute enrofloxacin treatment in albino rats

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Abstract

Background: The current study was conducted at the department of veterinary pharmacology and toxicology to assess the impact of *Emblica officinalis* on oxidative stress produced by sub-acute enrofloxacin exposure in albino rats.

Methodology: Albino rats weighing 200-300 gram were given enrofloxacin and *Emblica officinalis* orally through gavage needle, and blood samples were taken via capillary tube on the 28th day of the experiment and analysed using laboratory procedures.

Result: Sub-acute treatment of Enrofloxacin at doses of 5 mg/kg b.wt. and 10 mg/kg b.wt. generated oxidative stress in rats, as evidenced by substantial changes in oxidative parameters, and affected liver and kidney functions, as seen by changes in biochemical markers of liver and kidney function. *Emblica officinalis* aqueous extract (200 mg/kg b. wt.) effectively reduces oxidative stress and changes in hepatic and renal function caused by enrofloxacin.

Conclusion: *Emblica officinalis* (Amla) has antioxidant properties and the ability to reduce oxidative stress caused by enrofloxacin in albino rats following subacute treatment.

Keywords: Enrofloxacin, Emblica officinalis, albino rats, oxidative stress

Introduction

Enrofloxacin, a 6-fluoroquinolone, is one of the most widely used antimicrobials in veterinary practice. It is a bactericidal drug with a broad spectrum antibacterial activity against a range of bacteria, including the strains resistant to other antimicrobial agents. The mechanism of action of enrofloxacin consists of inhibition of bacterial DNA gyrase, which plays basic role in the process of DNA replication, leading to inhibition of synthesis of bacterial proteins. The metabolism of enrofloxacin takes place in the liver through the N-oxidation, N-dealkylation and de-ethylation processes. Enrofloxacin is mainly excreted through urine and bile. Fluoroquinolone antibiotics may cause adverse effects such as chondrotoxicity, renal damage, retinal damage, dys-glycemia, cardiac arrhythmia and even tendon rupture. These side effects of fluoroquinolones are associated with reduced collagen synthesis and induced oxidative stress (Elbe *et al.*, 2015) [12]. Induction of oxidative stress has been reported with the administration of ofloxacin, ciprofloxacin, levofloxacin, gatifloxacin and enrofloxacin (Ibrahim and Yarsan, 2011) [19]. There is evidence that free radical formation plays an important role in fluoroquinolone induced cartilage defect and photo toxicity (Gurbay *et al.*, 2006) [17].

Herbal products have a special place in the world of pharmaceuticals. Interests in the medicine of plant origin are spreading world-wide because of their safety, efficacy and cost effectiveness and negligible side effects. A number of plants have been mentioned in ayurveda for curing hepatic and renal diseases. The world health organization found that 80 percent of the world population depends on medicinal plant for their health care needs, and more than 30% of the pharmaceutical preparations are based on plants (Shinwari and Khan, 1998) [39]. *Emblica officinalis*, commonly known as Indian gooseberry or Amla, belonging to family Euphorbiaceae, is a main herbal drug utilized in unani and ayurvedic system of medicine (Bhandari and Kamdod, 2012) [6].

Material and Method

The suggested study was carried out on healthy albino rats weighing 150-200 g in the Department of Veterinary Pharmacology and Toxicology. The Institutional Animal Ethical Committee (IAEC) of the College of Veterinary Science and Animal Husbandry, Nanaji Deshmukh Veterinary Science University (NDVSU), Jabalpur, gave its approval to the study. For acclimatisation, the rats were maintained in laboratory conditions for 7 days prior to the start of the experiment. The rats were kept in colony cages under standard management and given standard meal and water ad -libitum in order to maintain good sanitary conditions.

Drugs

Fresh fruit of *Emblica officinalis* (Amla) was collected from Department of Botany, Jawaharlal Nehru Krishi Vishwa Vidyalaya, Jabalpur (M.P). Enrofloxacin was administered orally for subacute exposure.

Emblica officinalis aqueous extract preparation

To make powder, the fruits of *Emblica officinalis* were dried and crushed in a combination and grinder. Cold extraction was used to make the aqueous extract of *Emblica officinalis* (Shukla, 2006) [40]. The needed amount of *Emblica officinalis* fruit powder was weighed, steeped in distilled water, and kept at room temperature overnight. Filtration with filter paper yielded the cold aqueous extract.

Experimental Design

A total of 30 rats were randomly divided into five groups, each group with six rats. The effects of sub-acute enrofloxacin exposure on oxidative stress and organ damage, as well as the ameliorative potential of *Emblica officinalis* against these effects, were studied in five groups of rats, ranging from group I to group V. The trial took place over the course of 28 days.

Group	Treatment
I	Control
II	Enrofloxacin @ 5 mg/kg b. wt. once daily, orally for 28 days.
III	Enrofloxacin @ 10 mg/kg b. wt. once daily, orally for 28 days.
IV	Enrofloxacin @ 5 mg/kg b. wt. along with aqueous extract of <i>Emblica officinalis</i> @ 200mg/kg b.wt. once daily, orally for 28 days.
V	Enrofloxacin @ 10 mg/kg b. wt. along with aqueous extract of Emblica officinalis @ 200 mg/kg b. wt. once daily, orally for 28 days.

Collection of blood sample: Blood was taken from the retro-orbital plexus using a capillary tube on days 0 and 28 as reported by Archar and Riley (1981) ^[4]. Biochemical and oxidative stress parameters were studied using blood obtained in heparinized vials.

Biochemical studies

For biochemical analysis, plasma was extracted from heparinized blood samples and kept at 4 °C. The biochemical indicators of liver and kidney function were determined using a semi-automated analyzer and commercially available ERBA kits from Transasia Bio-Medicals Ltd., Daman.

Oxidative stress indicators evaluation

The samples were centrifuged at 2000 rpm for 15 minutes after blood collection to separate plasma. The layer of white blood cells above the packed erythrocytes was discarded. Erythrocyte pellet was washed three times with 0.15 M NaCl, diluted (33%) in phosphate buffer saline (mM: NaCl, 136.9, KCl, 2.68; KH₂·PO₄, 1.47; and Na₂·HPO₄, 6.62; pH 7.4) and kept at 4°C until further analysis. The 33% packed erythrocytes were used for the estimation of LPO, GSH, Glutathione reductase, Catalase and Superoxide dismutase activity by using Helios double beam spectrophotometer. LPO and GSH were measured on the day of blood collection (Prins and Loos, 1969) [33].

Lipid peroxidation

Lipid peroxidation (LPO) was estimated in 33% packed cell erythrocytes prepared in phosphate buffer saline of pH 7.4. Membrane peroxidative damage in erythrocytes was determined in terms of malondialdehyde (MDA) production as method suggested by Rehman (1984) [36].

Reduced glutathione (GSH)

200 µL whole blood was used for the estimation of blood

glutathione (GSH). GSH was estimated by the 5, 5' dithiobis (2-nitrobenzoic acid) (DNTB) method suggested by (Habig *et al.*, 1974) [18].

Superoxide dismutase

The activity of superoxide dismutase (SOD) in the blood supernatant was measured using the Madesh and Balasubramanian technique (1998). The suppression of superoxide dependent reduction of tetrazolium dye (MTT) [3-(4-5 dimethyl thiazol- 2-4) 2, 5-diphenyl tetrazolium bromide] in a calorimetric experiment. One unit of SOD was responsible for a 50% drop in MTT reduction rate. (Marklund and Marklund, 1974) [26].

Catalase

Catalase activity in erythrocytes was assayed by the spectrophotometric method of Aebi (1983) [1].

Reagents: Phosphate buffer saline, Haemolysate (10 per cent), Hydrogen peroxide (10 mM)

Statistical analysis

Means and standard error were obtained as per standard procedure. Parameters were analyzed by using the method of complete randomized design with seven treatments allotted to groups of six animals each. The difference between treatments was tested statistically for their significance (Snedecor and Cochran, 1994) [42].

Biochemical Studies ALT (SGPT) and AST (SGOT)

The concentration of ALT in control was $79.75 \pm 7.50 \text{IU/L}$. Enrofloxacin, significantly increased the concentration of ALT and this increase was $91.38 \pm 0.22 \text{U/L}$ at the dose rate of 5 mg/kg b.wt., orally for 28 days. In higher doses i.e.10 mg/kg b.wt. orally for 28 days, enrofloxacin further increased the concentration of ALT to 106.32 ± 0.10 A

significant amelioration by *E officinalis* was observed in group IV, where rats were treated with enrofloxacin at the dose rate of 5 mg/kg b.wt. and *Emblica officinalis* @ 200 mg/kg b.wt., orally for 28 days. In this group the mean values of ALT were $78.55\pm .14$ IU/L, which showed nonsignificant difference from control. This amelioration of *E. officinalis* was also observed, when enrofloxacin administered in higher doses i.e. 10 mg/kg b.wt. along with *Emblica officinalis* @ 200mg/kg b.wt., orally for 28 days. In this group the mean values of ALT were 71.92 ± 1.73 IU/L, which showed non-significant difference from control.

The concentration of AST in control was 159.98 ± 6.27 Enrofloxacin. significantly increased concentration of AST and this increase was 177.33 ± 0.49U/L at the dose rate of 5 mg/kg b.wt., orally for 28 days. In higher doses i.e.10 mg/kg b.wt., orally for 28 days, enrofloxacin further increased the concentration of AST to 180.53 ± 0.09 U/L. A significant amelioration by E. officinalis was observed in group IV, where rats were treated with enrofloxacin at the dose rate of 5 mg/kg b.wt. and Emblica officinalis at the rate 200 mg/kg b.wt., orally for 28 days. In this group the mean values of AST were 165.33 ± 3.22 IU/L, which showed non-significant difference from control. This amelioration of Emblica officinalis was also observed, when enrofloxacin administered in higher doses i.e. 10 mg/kg b.wt. along with Emblica officinalis at the rate of 200mg/kg b.wt., orally for 28 days. In this group the mean values of AST were 165.00 ± 3.71 IU/L, which showed non- significant difference from control.

The current study found that following subacute enrofloxacin exposure, serum ALT and AST levels were raised, which agrees with Guo *et al.* (2018) ^[16] findings. The liver enzyme markers ALT and AST are utilised to detect enrofloxacin-induced hepatic alterations in albino rats (Jiang *et al.*, 2014) ^[21]. In carbon tetrachloride-induced liver injury, Gulati *et al.* (1995) ^[15] found that Emblica officinalis demonstrated a substantial reduction in increased levels of SGPT and SGOT. These findings are consistent with the findings of the current study.

GGT (Gamma Glutamyl Transferase)

The concentration of GGT (Gamma glutamyl transferase) in control was 7.07 ± 0.03 IU/L. Enrofloxacin, significantly increased the concentration of GGT and this increase was 7.40 ± 0.04 IU/L after the subacute exposure of enrofloxacin at the dose rate of 5 mg/kg b.wt., orally for 28 days. In higher doses i.e.10 mg/kg b. wt., orally for 28 days enrofloxacin further increased the concentration of GGT to 7.70 ± 0.05 IU/L. A significant amelioration by *Emblica* officinalis was observed in group IV, where rats were treated with enrofloxacin at the dose rate of 5 mg/kg b.wt. and Emblica officinalis at the rate 200 mg/kg b.wt., orally for 28 days. In this group the mean values of GGT were 7.20 ± 0.04 IU/L, which showed non-significant difference from control. This amelioration of E. officinalis was also observed, when enrofloxacin administered in higher doses i.e. 10 mg/kg b.wt., along with Emblica officinalis at the rate of 200 mg/kg b.wt., orally for 28 days. In this group the mean values of GGT were 7.28± 0.04 IU/L, which showed non- significant difference from control.

Serum GGT is a liver enzyme that is sensitive, specific, and suggestive of liver function. According to Sureshkumar *et al.* (2013) [43], enrofloxacin enhanced the amount of the

enzyme GGT in blood following sub-acute dosing. Nagaraj *et al.* (2007) ^[30] found that an aqueous extract of Emblica officinalis (Amla) reduced the raised level of GGT in rat liver after exposure to hexachloro-cyclohexane-induced cytotoxicity.

ALP (Alkaline Phosphatase)

The concentration of alkaline phosphatase in control was 449.50± 2.54. Enrofloxacin, significantly increased the concentration of ALP and this increase was 532.50 ± 0.96U/L after the subacute exposure of enrofloxacin at the dose rate of 5 mg/kg b.wt., orally for 28 days. In higher doses i.e.10 mg/kg b.wt., orally for 28 days, enrofloxacin further increased the concentration of ALP to 545.17 ± 3.71 IU/L. A significant amelioration by Emblica officinalis was observed in group IV, where rats were treated with enrofloxacin at the dose rate of 5 mg/kg b.wt. and Emblica officinalis @ 200mg/kg b.wt., orally for 28 days. In this group the mean values of ALP were 450.17 ± 2.69 IU/L, which showed non-significant difference from control. This amelioration of E. officinalis was also observed, when enrofloxacin administered in higher doses i.e. 10 mg/kg b.wt. along with Emblica officinalis @ 200mg/kg b.wt., orally for 28 days. in this group the mean values of ALP were 461.33 ± 0.7 U/L, which showed non-significant difference from control.

Alkaline phosphatase is used to detect liver disease and is also indicative enzyme for liver function. Elkholy *et al.* (2009) [13] showed that administration of enrofloxacin @ 10 mg /kg body weight daily for five successive days caused elevation of serum ALP when given orally. Davoren and Mainstone (1993) [10] and Moustafa *et al.* (1998) [28] showed that oral administration of enrofloxacin for 15 consecutive days caused elevation in serum ALP level in dogs. The investigations by Reddy *et al.* (2009) [35], mentioned the protective role of *Emblica officinalis* on alcohol induced liver damage in respect of ALP level in albino rats.

Albumin and Bilirubin

The concentration of albumin in control was 4.05 ± 0.03 . Enrofloxacin, significantly increased the concentration of albumin and this increase was 4.33 ± 0.03 g/dl after the administration of enrofloxacin at the dose rate of 5 mg/kg b.wt., orally for 28 days. In higher doses i.e.10 mg/kg b.wt., orally for 28 days, enrofloxacin further increased the concentration of albumin to 4.50 ± 0.05 g/dl. A significant amelioration by Emblica officinalis was observed in group IV, where rats were treated with enrofloxacin at the dose rate of 5 mg/kg b.wt. and Emblica officinalis @ 200 mg/kg b.wt., orally for 28 days. In this group, the mean values of albumin were 4.10± 0.02 g/dl, which showed nonsignificant difference from control. This amelioration of Emblica officinalis was also observed, when enrofloxacin administered in higher doses i.e. 10 mg/kg b.wt. along with Emblica officinalis @ 200 mg/kg b.wt., orally for 28 days. In this group, the mean values of albumin were 4.15 ± 0.03 g/dl, which showed non- significant difference from control. In the present study the values of albumin were measured on day 28 of the experiment, the concentration of albumin was significantly higher in enrofloxacin treated group as compared to control group.

The concentration of Bilirubin in control was 0.107 ± 0.003 mg/dl. Enrofloxacin, significantly increased the concentration of bilirubin and this increase was $0.127\pm$

0.003 mg/dl after the administration of enrofloxacin at the dose rate of 5 mg/kg b.wt., orally for 28 days. In higher doses i.e.10 mg/kg b.wt., orally for 28 days, enrofloxacin further increased the concentration of bilirubin to 0.137± 0.002 mg/dl. A significant amelioration by Emblica officinalis was observed in group IV, where rats were treated with enrofloxacin at the dose rate of 5 mg/kg b.wt. and Emblica officinalis @ 200 mg/kg b.wt., orally for 28 days. In this group, the mean values of bilirubin were which 0.110 ± 0.004 mg/dl, showed non-significant difference from control. This amelioration of E. officinalis was also observed, when enrofloxacin administered in higher doses i.e. 10 mg/kg b.wt. along with Emblica officinalis @ 200 mg/kg b.wt., orally for 28 days. In this group, the mean values of bilirubin were 0.117 ± 0.002 mg/dl, which showed non- significant difference from control.

Enrofloxacin dosing resulted in a considerable rise in albumin and bilirubin in the current investigation. Enrofloxacin raised albumin and bilirubin levels following subacute treatment, according to Kock *et al.* (1987) ^[24]. *Emblica officinalis* (Amla), which is rich in vitamin C, gallic acid, flavonoids, and tannins, protects against enrofloxacin-induced hepatotoxicity and lowers liver enzyme concentrations (Reddy *et al.* 2009) ^[35].

Biochemical markers of kidney function Creatinine and blood urea nitrogen

The concentration of creatinine in control was 0.640±0.021 Enrofloxacin, significantly increased concentration of creatinine and this increase was $0.847 \pm$ 0.013 mg/dl after the administration of enrofloxacin at the dose rate of 5 mg/kg b.wt. In higher doses i.e.10 mg/kg b.wt enrofloxacin further increased the concentration of creatinine to 1.052±0.023 mg/dl. In group IV, where rats were treated with enrofloxacin at the dose rate of 5 mg/kg b.wt., and Emblica officinalis at the rate of 200 mg/kg b.wt. orally for 28 days, the mean values of creatinine were 0.840 ±0.013 mg/dl, which was significantly lower than enrofloxacin (10 mg/kg) treated group. A significant amelioration by Emblica officinalis was observed, when enrofloxacin administered in higher doses i.e. 10 mg/kg b.wt. along with Emblica officinalis in group V. In this group the mean values of creatinine were 0.913±0.005 mg/dl, which were significantly lower than enrofloxacin alone (10 mg/kg) treated group.

The concentration of Blood urea nitrogen in control was 14.35± 0.23 mg/dl. Enrofloxacin, at the dose rate of 5 mg/kg b.wt., orally for 28 days did not affect the BUN level in serum significantly and the mean value was 14.62± 0.03. However, enrofloxacin in higher doses i.e.10 mg/kg b.wt., orally for 28 days, significantly increased the concentration of BUN to 23.84± 0.02 mg/dl. In group IV, where rats were treated with enrofloxacin at the dose rate of 5 mg/kg b.wt. and Emblica officinalis @ 200 mg/kg b.wt., orally for 28 days, the mean value of BUN was 14.36± 0.02 mg/dl, which showed non-significant difference from control. A significant amelioration by E. officinalis was observed in group V where rats were treated with enrofloxacin at the dose rate of 10 mg/kg b.wt. and Emblica officinalis @ 200 mg/kg b.wt., orally for 28 days. In this group the mean value of BUN was 19.12± 0.01 mg/dl, which was significantly lower than enrofloxacin (10 mg) treated group.

According to Khan and Rampal (2014) [22], sub-acute

enrofloxacin exposure raised creatinine and BUN concentrations. In hyperammonemic patients, Krishnaveri *et al.* (2010) ^[45] found that Emblica officinalis restored blood urea nitrogen and creatinine levels by boosting antioxidant status. The conclusions of this inquiry corroborate those of the mentioned researchers.

Oxidative stress indices Lipid peroxidation (MDA)

Lipid peroxidation was calculated in terms of nM MDA/gm of blood on day 28 of experiment in albino rats. The concentration of LPO in control was 4.63±0.02 nM MDA/gm of blood, However, enrofloxacin at dose of 5 mg/kg b.wt and 10 mg/kg b.wt. significantly increased the concentration of LPO and the mean values were 5.20 ±0.03 and 5.78±0.03 nM MDA /gm of blood, respectively. A significant amelioration of enrofloxacin induced oxidative stress was shown by *Emblica officinalis* and was observed by reduced LPO concentration in group IV and V with mean values 4.17 ±0.04 and 4.83±0.03 respectively.

Sarban *et al.* (2005) [38] discovered a rise in MDA levels in the plasma after taking enrofloxacin, as evidenced by the appearance of lipid hydroperoxide. In this study, oral administration of *Emblica officinalis* aqueous extract at the time of enrofloxacin exposure significantly reduced the degree of lipid peroxidation. This might be due to the existence of radical scavengers with antioxidant properties that can reduce the generation of peroxides, hydroxyl radicals, and superoxide radicals. Our findings are consistent with those of Bast *et al.* (1991) [5], Bhattacharya *et al.* (1999) [7], and Khopde *et al.* (2001) [23], who all found that aqueous extract of Emblica officinalis significantly reduced LPO content in albino rats, indicating free radical scavenging action.

SOD (Superoxide Dismutase)

Superoxide dismutase was calculated in terms of U/g of Hb on day 28 of experiment in albino rats. The concentration of SOD in control was 1.22±0.01 U/g of Hb, however enrofloxacin at dose of 5 mg/kg b.wt and 10 mg/kg b.wt. significantly decreased the concentration of SOD and the mean values were 0.88 ± 0.01 and 0.84 ± 0.02 U/g respectively. A significant amelioration of enrofloxacin induced oxidative stress was shown by Emblica officinalis and was observed by elevated SOD concentration in group IV and V with mean values 1.12±0.03 and 1.14±0.04 respectively. SOD activity contributes to increasing the level of superoxide radicals, thus increased oxidative stress. Sureshkumar et al. (2013) [43] indicated that enrofloxacin administration reduced superoxide dismutase (SOD). The aqueous extract of Emblica officinalis (Amla) was found to increase the activity of antioxidant enzymes. Bhattacharya et al. (2002) [8] and Rajak et al. (2004) [34] reported that an aqueous extract of Emblica officinalis boosted the activity of the antioxidant enzyme SOD in rats, and these findings are consistent with those of the current study.

GSH (Reduced Glutathione)

Reduced glutathione was calculated in terms of $\mu mol\ /ml$ of blood on day 28 of experiment in albino rats. The concentration of GSH in control was 340.67±0.21 $\mu mol\ /ml$ of blood, However, enrofloxacin at dose of 5 mg/kg b.wt and 10 mg/kg b.wt. significantly decreased the concentration of GSH and the mean values were

332.57 \pm 0.24 and 330.42 \pm 0.44 µmol /ml of blood respectively. A significant amelioration of enrofloxacin induced oxidative stress was shown by *Emblica officinalis* and was observed by increased GSH concentration in group IV and V with mean values 334.33 \pm 0.17 and 335.42 \pm 0.35 µmol /ml of blood respectively.

According to Cerreras et al., (2005) [9], metabolism of enrofloxacin residues generates free radicals and contributes to an increase in oxidative stress, resulting in cellular enzyme inhibition owing to glutathione peroxidase decrease (GSHPx). In the current study, GSH concentration was considerably lower following enrofloxacin compared to the control group. In agreement with the fact that GSH concentration was significantly lower after enrofloxacin treatment compared to the control group in the current study (Sai ram et al., 2002 [37], Alia et al., 2006 [2], Anila et al., 2006) [2], Emblica officinalis (Amla) normalised glutathione peroxidase levels by increasing antioxidant status in the oxidative state. When the findings of the current inquiry are considered collectively, it is clear that they support the conclusions of the previous researchers.

Catalase

Catalase was calculated in terms of µmol H₂O₂ decomposed/min/gm Hb on day 28 of experiment in albino rats. The concentration of CAT in control was 235.08±0.38 umol H₂O₂ decomposed. However, enrofloxacin at dose of 5 mg/kg b.wt and 10 mg/kg b.wt. significantly decreased the concentration of CAT and the mean values were 157.83 ± 0.48 and 131.57 ± 0.20 µmol H_2O_2 decomposed, respectively. A significant amelioration of enrofloxacin induced oxidative stress was shown by Emblica officinalis and was observed by increased CAT concentration in group IV and V with mean values of 231.50±0.34 and 209.92±0.24, respectively. According to Yazar and Tras (2001) [44], the metabolism of enrofloxacin residues generates free radicals and contributes to an increase in oxidative stress, resulting in cellular enzyme inhibition owing to a decrease in catalase (CAT). The current study's findings are consistent with Yazar and Tras (2001) [44], since CAT concentration was considerably lower in the enrofloxacin-treated group compared to the control group. In arsenic-induced oxidative stress, Singh et al. (2014) [41] found that a crude aqueous extract of Emblica officinalis (Amla) enhanced catalase levels. When compared to data from rats treated with arsenic alone, amla co-treatment boosted activity by 77%, suggesting a protective effect against oxidative damage.

Conclusions

It is conclude that enrofloxacin at doses of 5 mg/kg body wt. and 10 mg/kg body wt., administered orally daily for 28 days in albino rats, induced liver and kidney impairment as indicated by increased concentrations of biochemical markers of liver and kidney function, and at doses of 5 mg/kg body wt. and 10 mg/kg body wt., enrofloxacin significantly induced oxidative stress after subacute exposure. Aqueous extract of *Emblica officinalis* at the rate of 200 mg/kg b. wt., orally, daily for 28 days dramatically improved biochemical indicators of liver and kidney functioning, as well as oxidative stress indices.

References

1. Aebi HE. Catalase, In: H.U. Bergmeyer, J. Berg-meyer and M. GraBI, Eds., Methods of Enzymatic Analysis,

- 3rd Edition, Verlag Chemie, Weinheim, b, 1983, 273-286pp.
- Alia M, Mateos R, Ramos S, Lecumberri E, Bravo L, Goya L. Influence of quercetin on growth and antioxidant defense system of a human hepatoma cell line (HepG2). Europian Journal of Nutrure. 2006;45:19-28.
- 3. Anila L, Vijayalakshmi NR. Antioxidant action of flavonoids from *Mangifera indica* and *Emblica officinalis* in hypercholesterolemic rats. Journal of Food Chemistry. 2003;83(4):569-574.
- 4. Archar RK, Riley J. Standardized method of bleeding rats. Laboratory Animals. 1981;15:25-28.
- 5. Bast A, Maenon GR, Doelman GJ. Antioxidant activity of plants herb. American Journal of Medicine. 1991:91:251-254.
- 6. Bhandari PR, Kamdod MA. *Emblica officinalis* (Amla): a review of potential therapeutic applications. International Journal of Green Pharmacy. 2012;6(4):257-269.
- 7. Bhattacharya A, Chattergee A, Ghosal S, Bhattacharya SK. Antioxidant activity of active tannoids principles of *Emblica officinalis* (amla). Indian Journal of Experimental Biology. 1999;37:676-80.
- 8. Bhattacharya SK, Bhattacharya A, Sairam K, Ghosal S. Effect of bioactive tannoids principles of *Emblica officinalis* on ischemia reperfusion- induced oxidative stress in rat heart. Journal of Phytomedicine, 2002;9:171-174.
- Carreras I, Castellari M, Valero A, Antonio J, Sarraga C. Influence of enrofloxacin administration on the proteolytic and antioxidant enzyme activities of raw and cooked turkey products. Journal of Food Science and Agriculture. 2005;85:2407-2412.
- 10. Davoren P, Mainstone K. Norfloxacin induced hepatitis. Medical Journal of Australia. 1993;159(6):423-426.
- 11. Devran C, Burk D, Hotice F, Kurtulus P. Effect of enrofloxacin on joint fluid/blood status and organ damage markers. Journal of Annual Research and Review In Biology. 2018;25(3):1-7.
- 12. Elbe H, Dogan Z, Taslidere E, Cetin A, Turkoz Y. Beneficial effect of quercetin on renal injury and oxidative stress induced by ciprofloxacin in rats. Journal of Human and Experimental Toxicology, 2015;35(3):276-281.
- 13. Elkholy H, Elkomy A, Awaidat S, Elmajdoub AA. Tissue and egg residues and adverse effect of two oral enrofloxacin preparations; Baytril and Enrotryl. Journal of Global Veterinaria. 2009;3(5):363-368.
- 14. Girish C, Pandey S. Indian herbal medicines in the treatment of liver diseases: Problems and promises. Fundaments of Clinical Pharmacology. 2012;26:180-190.
- 15. Gulati RK, Agarwal S, Sharma SS. Hepatoprotective studies on *Phyllanthus emblica* Linn and quercetin. Indian Journal of Experimental Biology. 1995;33(4):261-268.
- 16. Guo J, Zhang D, Imal M. Advance in the research on the mechanism of to nifying kidney herbs that protect the articular cartilage. Journal of Traditional Chinese Medicine. 2018;28:2515-2518.
- 17. Gurbay A, Gonthier B, Singorini N, Barret L, Favier A, Hincal F. Ciprofloxacin induced DNA damage in

- primary culture of rat astrocyte and protection by Vitamin E. Journal of Neurotoxicoloy. 2006;27(1):6-10.
- 18. Habig WH, Pabst MJ, Jakoby WB. Glutathione-S-transferase: the first enzymatic step in mercapturic acid formation. Journal of Biological Chemistry, 1974;249:7130-7139.
- 19. Ibrahim IG, Yarsan E. Enrofloxacin drug induced reactive oxygen species. Journal of Research Opinion in Animal and Veterinary Sciences. 2011;1(8):489-491.
- 20. Jeena K, Joy L, Kuttan R. Effect of *Emblica officinalis*, Phyllanthus amarus and Picrorrhizakurroa on N-nitrosodiethylamine induced hepatocarcinogenesis, Cancer Letter. 1999;136:11-16.
- 21. Jiang L, Wang W, Luo L. Blood concentration and histological toxicology of different dosages of florfenicol in GIFT Nile tilapia. Journal of Shanghai Ocean University. 2014;23:90-94.
- 22. Khan AM, Rampal S. Effect of repeated oral administration of levofloxacin, enrofloxacin, and meloxicam on antioxidant parameters and lipid peroxidation in rabbits. Human and Experimental Toxicology. 2014;36(1):216-218.
- 23. Khopde SM, Priyadarsini KI, Mohan M, Banabaliker MM. Charecterising the antioxidant activity of amla (*Phyllanthus emblica*) extract. Journal of Current Science. 2001;8(1):185-190.
- 24. Kock MD, Jessup D, Clark R, Frantic C. Effects of capture on biological parameters in free-ranging bighorn sheep (*Ovis canadensis*): Evaluation of dropnet, drive-net, chemical immobilization and the netgun. Journal of Wildlife Diseases. 1987;23:641-651.
- 25. Islam SJ, Deka C, Sonowal PJ. Treatment and management of lumpy skin disease in cow: A case report. Int. J. Vet. Sci. Anim. Husb. 2021;6(2):26-27.
- 26. Marklund S, Marklund G. Involvement of the superoxide anion radical in the autoxidation of pyrogallol and a convenient assay for superoxide dismutase. European Journal of Biochemistry. 1974;47:469-474
- Mehta S, Singh RK, Jaiswal D, Rai P, Watal G. Antidiabetic activity of Emblica officinalis in animal models. Pharmaceutical biology. 2009;47(11):1050-1055.
- 28. Moustafa A, Baset I, Mashad A, Tantawy A. Pathological studies on enrofloxacin and its residues in broiler chickens. Egyptian Journal of Comparative Pathology. 1998;11:57-66.
- 29. Nagraj R, Deb U, Bhaskar S, Prasad G, Rao PV. Hepatoprotective efficacy of certain flavonoids against microcystin induced toxicity in mice, Environmental Toxicology. 2007;22:472-479.
- 30. Pandey GP. Studies on phytochemistry and acute toxicity of Livol and its some ingredients. Indian Journal of Indigenous Medicine. 2011;6:55-64.
- 31. Patel SS, Goyal RK. *Emblica officinalis* Geart: A Comprehensive Review on Phytochemistry, Pharmacology and Ethnomedicinal Uses. *Research* Journal of Medicinal Plant. 2012;6:6-16.
- 32. Pramyothin P, Patcharavadee S, Somlak P, Chaiyo C. The protective effects of *Phyllanthus emblica* Linn. extract on ethanol induced rat hepatic injury, Journal of Ethno-pharmacology. 2006;107:361-364.

- 33. Prins HK, Loos. Biochemical methods in red cell genetics. Yunis, J.J. Glutathion. Publ. Academic press. NY, 1969, 115-137pp.
- 34. Rajak S, Banerjee SK, Sood S, Dinda AK, Gupta YK, Gupta SK, *et al. Emblica officinalis* causes myocardial adaptation andprotects against oxidative stress in ischemic-reperfusion injury in rats. Journal of Phytochemistry. 2004;18:54-60.
- 35. Reddy VD, Padmavathi P, Paramahamsa M, Varadacharyulu N. Amelioration of alcohol induced oxidative stress by Emblica officinalis (amla) in rats. Indian Journal of Biochemistry and Biophysics. 2009;47:20-25.
- 36. Rehman SU. Lead induced regional lipid peroxidation in brain. Toxicology letters. 1984;21(3):333-337.
- 37. Sai Ram M, Neetu D, Deepti P. Cytoprotective activity of Amla (*Emblica officinalis*) against chromium (VI) induced oxidative injury in murine macrophages. Journal of Pharmacological Research. 2002;17(4):430-433.
- 38. Sarban A, Sezgin A, Koenig B, Mithat Y, Augur A, Isikan E. Plasma total antioxidant capacity, lipid peroxidation, and erythrocyte antioxidant enzyme activities in patients with rheumatoid arthritis and osteoarthritis. Journal of Clinical Biochemistry, 2005;38:981-986.
- 39. Shinwari MI, Khan P. Indigenous use of medicinal trees and shrubs of Margalla Hills National Park, Islamabad. Pakistani Journal of Forest. 1998;48:63-90.
- 40. Shukla D. Studies on pharmacological action of *Boerhaavia diffusa* with special reference to its antibacterial activity in rats. M.V.Sc. & A.H. thesis (Pharmacology and Toxicology), Nanaji Deshmukh Veterinary Science University, Jabalpur. 2006.
- 41. Singh M, Yadav R, Yogeshwar S, Shukla S, Sanjay S. Efficacy of crude extract of *Emblica officinalis* (amla) in arsenic-induced oxidative damage and apoptosis in splenocytes of mice. Toxicology Ietter. 2014;21(1):8-17.
- 42. Snedecor GW, Cochran WG. Statistical Methods, 8th Edn., The lowa State College Press, Inc. USA. 1994, 950pp.
- 43. Sureshkumar V, Sarathchandra G, Ramesh J. Veterinary pharmacovigilance evaluation on impact of enrofloxacin administration on antioxidant status in broiler chicken. International Journal of Current Microbiological Science. 2013;2:335-341.
- 44. Yazar E, Tras B. Effect of fluoroquinolone antibiotic on hepatic superoxide dismutase and glutathione peroxidase activates in healthy and experimentally induced peritonitis mice. Revue de medecineveterinaire, 2001;15:200-235.
- 45. Krishnaveni M, Swathi A, Mirunalini S. Therapeutic potential of Phyllanthus emblica (amla): the ayurvedic wonder. Journal of Basic Clinical Physiology and Pharmacology. 2010;21:93-105.